

SYNTAXONOMY: A SOURCE OF USEFUL FUZZY SETS FOR ENVIRONMENTAL ANALYSIS?

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Abstract. The use of vegetation types defined in syntaxonomical research is proposed for environmental analysis on the basis of fuzzy set theory. An example with data of a relatively simple environmental system is discussed.

Introduction

Syntaxonomy may be considered as a source of fuzzy sets, being one of its aims the definition of vegetation types at different hierarchical levels (see Dierschke 1981). In fact each vegetation type consists of two sets of objects: one of species S found in the communities and the other of relevés R on which the type is based. Since the same species may occur in many different types and the relevés are assigned to types on the basis of the presence of species, or on the basis of a similarity level (polythetic assignment, Dale 1977) it would appear to be justifiable to attach a degree of membership to each species in S :

$$F(S) = \{s, f(s)\}$$

and to each relevé in R :

$$F(R) = \{r, f(r)\}$$

where s and r are respectively species and relevés and f is the degree of membership in a type.

In fuzzy set theory (Zimmerman 1985); introduced into plant ecology explicitly by Marsili-Libelli (1986), Roberts (1986) and Feoli and Zuccarello (1986), the degree of membership of the elements to a set ranges continuously between 0 and 1; while in traditional set theory it is either 0 or 1. In fact, the description of a vegetation type by the set of species and by the set of relevés is rarely a yes or no proposition, *i.e.*, categorically distinct, therefore the application of fuzzy set theory to community ecology seems particularly appropriate (Dale 1977).

In this paper the fuzzy set theory is applied to a relatively simple ecological system in order to demonstrate how the syntaxa can be used to explain trends of variations in relation to gradients and dynamics. Environmental fuzzy sets (EFS) are defined by chemical-physical data and used for ordering relevés of plant communities and species.

Data

A relatively simple syndynamic scheme of Giacomini and Pignatti (1955) representing the relationships between four vegetation types provided the data. The scheme is presented in Fig. 1. The relevés corresponding to the types are presented in Table 1 (matrix X with 69 rows and 50 columns, rows = species, columns = relevés). In this table the species of the two syntaxonomical classes: *Salicetea herbaceae* (including *Polytrichetum* and *Salicetum*) and *Caricetea curvulae* (including *Hygrocurvuletum*) are indicated by symbols, since only these species will be considered in the ordination scatter-plots. The average values for the measured environmental factors are presented in Table 2 (matrix E with 6 rows and 4 columns, rows = variables, columns = vegetation types). The syndynamic scheme on which the analysis is based has been simplified (p. 100, Fig. 11 in Giacomini and Pignatti 1955). *Polytrichetum* has been taken as a single type and *Caricetum fuscae* has been omitted (it shows only three species in common with the other types). The scheme of Fig. 1 represents a syndynamic series and reflects a gradient in which snow depth and snow persistence are the major controlling environmental factors. These both decrease from *Polytrichetum* to *Hygrocurvuletum*. For computations the Braun-Blanquet scale has been transformed according to van der Maarel (1979).

Method

The degree of membership of relevés in the types have been computed following the procedure suggested by Feoli and Zuccarello (1986). To obtain the environmental fuzzy sets (EFS) the steps include:

1) The relevés in Table 1 are classified by sum of squares clustering based on geodesic distance (Orlói 1978). This is comparable to the visual classification of Giacomini and Pignatti (1955).

2) Centroids are computed for each vegetation type according to Table 1 (matrix X 69×50). The centroids constitute the column vectors of the resulting matrix C (69×4).

| Community type n. | | 111111111111 | 222222222222 | 33333333333333333333 | 444444 |
|-------------------|---|-------------------|--|----------------------|--------|
| Relevé n. | | 11 11111111222222 | 22222333333333334444 | 444445 | |
| | | 12345678901 | 2345678901234 | 56789012345678901234 | 567890 |
| S1 | <i>Polytrichum sexangulare</i> | ++r54555543 | 22222211.+.+. .12+. .1+.+.r.111 | | |
| S2 | <i>Anthelja juratzkana</i> | 443.+.1++++ |3.+.r. | | |
| S3 | <i>Webera commutata</i> | 2++++.++++ | | | |
| S4 | <i>Webera carinata</i> | .+.+.+.+ | | | |
| S5 | <i>Kiaeria starkei</i> | ...+.+.+ |+.+.+.+.+.+ | | |
| S6 | <i>Pleuroclada albescens</i> | ...+.+.+ | | | |
| S7 | <i>Gymnomitrium varians</i> | ..2..... | | | |
| S8 | <i>Dicranum falcatum</i> | .2..... | | | |
| S9 | <i>Arenaria biflora</i> | 1++2+2+++ | +++1.1+.+. .1.11+++1.+. | | |
| S10 | <i>Gnaphalium subinum</i> | +++1.12+++ | 222+232+21..2 122112+2212112+1+1 |+ | |
| S11 | <i>Salix herbacea</i> | 23.+.+.+.+ | 5455554444544 ...12+...3. | ++2++1 | |
| S12 | <i>Soldanella pusilla</i> | .+.+.+.+.+ | +.+.+.+.+.+. +2+1+31+3+111112111+ | | |
| S13 | <i>Sibbaldia procumbens</i> | ...+.+.+.+.+ | ...+.+.+.+.+.+. +11..2+1 | | |
| S14 | <i>Alchemilla pentaphylla</i> | ...+.+.+.+.+ | ...21 55555535445535545454 | | |
| S15 | <i>Veronica alpina</i> | +.++++112++ | 1+.1+.1+.+.+. +1+.+.+.+.+ | | |
| S16 | <i>Gentiana bavarica</i> | ...+++11+ |+.+.+.+.+.+ | | |
| S17 | <i>Sedum alpestre</i> | +.+.+.+.+.+ | 11+++11.+.+.+.+.+.+ | | |
| S18 | <i>Sagina saginoides</i> | +.1.++ |+.+.+.+.+. 1.111.+.+.+.+.+ | | |
| S19 | <i>Cerastium cerastioides</i> | +1.+.+.+ | 1+2.1+.1+.+. .1.+.+.+.+.+ | | |
| . | <i>Chrysanthemum alpinum</i> | +++1+22122 | +1+12+1+1+2. .2++12+1++1+++1+++. | 1+1++ | |
| . | <i>Poa alpina</i> var. <i>minor</i> | +++++.1+++ | +1+12+1+1+21 2+12121121+31112121 | | |
| . | <i>Taraxacum alpinum</i> | +.+.+.11+1 | +++1+++1.+.+.+.+.+.+ | | |
| . | <i>Poa supina</i> | ...+.+.1+.+ | | | |
| . | <i>Ranunculus glacialis</i> | ...+.+.+.1 | | | |
| C1 | <i>Carex curvula</i> | +.+.+.+.+.+ | ...+.+.+.+.+. +1+.+.1+1++1.2333444 | 344555 | |
| . | <i>Cardamine alpina</i> | +.1.+ | 1+.+.+.+.+.+.+.+.+.+.+.+.+ | | |
| C2 | <i>Agrostis rupestris</i> | ...+.+.+.+.+ | ...+.+.+.11..2 1+...3.+.+.2.+.+.+.1 | +.1.1 | |
| . | <i>Carex lachenalii</i> | ...+.+.+.+.+ | | | |
| . | <i>Stereocaulum coralloides</i> | ...+.+.+.32 | | | |
| . | <i>Ochrolechia</i> sp. | ...11++ | +.1+1+.1.+.+.+.+.+.+.+.+ | | |
| . | <i>Dicranoweisia crispula</i> | ...+.+.+.+.+ | | | |
| . | <i>Cladonia pyxidata</i> | ...+.+.2+.+.+ | ...+.+.+.+.+.+.+.+.+.+.+.+.+ | | |
| . | <i>Cladonia rangiferina</i> | ...+.+.+.+.+ | ...+.+.+.+.+.+.+.+.+.+.+.+.+ | +2.+++ | |
| . | <i>Polytrichum piliferum</i> | ...+.+.23 | ...+.+.+.+.4.+.+.+.+.+.1+11+.+ | ...+.+ | |
| . | <i>Pogonatum alpinum</i> | ...2..... | | | |
| C3 | <i>Leontodon pyrenaicus</i> | | ...1+++1.+++++++ | 1+1.1+ | |
| . | <i>Polygonum viviparum</i> | | ...+.+.+.+.+.+.+.+.+.+.+.+.+ | 11++21 | |
| C4 | <i>Phyteuma hemisphaericum</i> | | ...+.+.+.+.+.+.+.+.+.+.+.+.+ | +.2. | |
| C5 | <i>Ligusticum mutellina</i> | | ...12223+2222211 | 1...12 | |
| C6 | <i>Potentilla aurea</i> | | ...+.+.+.+.1+++2+1 | ...2. | |
| . | <i>Sieversia montana</i> | | ...+.+.+.+.+.+.+.+.+.+.+.+.+ | ...+.+ | |
| C7 | <i>Nardus stricta</i> | | ...+.+.+.+.+.+.+.+.+.+.+.+.+ | | |
| C8 | <i>Minuartia sedoides</i> | | ...+.+.+.1.+.+.+.+.+.+.+.+.+.+.+ | ...+.+ | |
| C9 | <i>Euphrasia minima</i> | | ...1+3+.1.1.1.+.+.+.+.+.+.+.+.+.+.+ | ...+.+ | |
| . | <i>Euphrasia drosocalyx</i> | | ...+.2.+.1+.1.+.+.+.+.+.+.+.+.+.+.+ | ...+.+ | |
| . | <i>Luzula spicata</i> | | ...2...2...2.1.+.11.+.+.+ | +++.+1 | |
| . | <i>Luzula lutea</i> | | ...+.+.+.+.+.+.+.+.+.+.+.+.+.+.+.+ | ...+.+ | |
| . | <i>Cardamine resedifolia</i> | | ...1.1.+.+.+.+.+.+.+.+.+.+.+.+ | | |
| . | <i>Polytrichum juniperinum</i> | | ...+.2.+.+.+.+.+.+.+.+.+.+.+.+.+.+.+.+ | ...+.+ | |
| . | <i>Cetraria islandica</i> | | ...+.+.1.+.+.+. 1..1+.1+++1.1++21.+ | +3++12 | |
| . | <i>Rhacomitrium ericoides</i> | | ...+.+.+.+.3.+.+.+.+.+.+.+.+.+.+.+ | | |
| C10 | <i>Senecio carniolicus</i> | | | ...+.+++ | |
| C11 | <i>Hieracium glanduliferum</i> | | | ...+. | |
| . | <i>Thamnochloa vermicularis</i> | | | ...+. | |
| . | <i>Alectoria ochroleuca</i> | | | ...+. | |
| . | <i>Cetraria cucullata</i> | | | ...+. | |
| C12 | <i>Veronica bellidioides</i> | | | ...+.+.2 | |
| C13 | <i>Festuca halleri</i> | | | ...+. | |
| C14 | <i>Silene acaulis</i> ssp. <i>exscapa</i> | | | ...+. | |
| . | <i>Poa alpina</i> | | | ...221+ | |
| . | <i>Oreocloa distica</i> | | | 331.41 | |
| . | <i>Primula oenensis</i> | | | ...+.1.++ | |
| . | <i>Avenastrum versicolor</i> | | | ...1.+.+.+ | |
| . | <i>Soldanella alpina</i> | | | ...+. | |
| . | <i>Erigeron uniflorus</i> | | | ...+.+. | |
| . | <i>Phyteuma globulariaefolium</i> | | | ...1. | |
| . | <i>Cetraria aculeata</i> | | | ...2.+.+.+ | |
| . | <i>Cetraria nivalis</i> | | | ...+++1.+.+ | |
| . | <i>Paraleucobryum enerve</i> | | | ...+. | |

Table 1. Vegetation relevés used for computations representing the 4 types in Fig. 1. Codes for species: S1-19 (Salicetea), C1-14 (Caricetea).

3) C is multiplied by X :

$$C' X = Y$$

C and X are normalized by rows and columns. Because of normalization any element $y(i, r)$ of $Y (4 \times 50)$ is cosine of the angle between a relevé r and the centroid of a type i , i.e. a measure of similarity.

The fuzziness of a relevé with respect to type i is computed by:

$$f(i, r) = y(i, r) - y(i \min) / (y(i \max) - y(i \min)).$$

4) Fuzziness of species with respect to vegetation types are computed by:

$$U = F X'$$

where F are normalized by columns and rows. X is normalized by rows. Also the values in $U (4 \times 69)$ are cosines and are transformed in order to obtain the fuzziness according to step 3).

5) Fuzziness of relevés with respect to environmental factors (environmental fuzzy sets, EFS) are computed by:

Table 2. Description of vegetation types by centroids of environmental variables.

| | TYPE | | | |
|--|-------|-------|-------|-------|
| | 1 | 2 | 3 | 4 |
| pH | 4.60 | 4.80 | 4.70 | 4.46 |
| CaCO ₃ % | 1.28 | 0.26 | 0.58 | 0.38 |
| P ₂ O ₅ tot.% | 1.44 | 1.43 | 1.83 | 1.73 |
| P ₂ O ₅ sol.mg/l | 0.05 | 2.50 | 3.10 | 5.03 |
| Org.m.% | 7.80 | 11.30 | 16.30 | 30.53 |
| H ₂ O% | 23.50 | 26.10 | 30.50 | 37.73 |
| number ob. | 4 | 3 | 13 | 3 |

$$A = E F$$

with E normalized by rows and F by columns. Also the elements in $A (6 \times 50)$ are cosines and are transformed to obtain measures of fuzziness to step 3).

6) Fuzziness of species with respect to environmental factors (EFS) are computed by:

$$Z = A X' \text{ or } Z = E U$$

with A normalized by columns and rows, X by rows, E by rows and U normalized by columns. Also the values in $Z (6 \times 69)$ are transformed as in step 3).

The row vectors of matrices F , U , A and Z can be used directly as ordination axes or by combining them two by two according to the anticommutative difference (Roberts 1986). For types i, h with respect to relevé r :

$$f(i/h)_r = (1 + (1 - f(h)_r)^2 - (1 - f(i)_r)^2) / 2.$$

The ordinations of relevés and species obtained by the same fuzzy sets may be superimposed (e.g., A on Z) to demonstrate the correspondence between species and relevés in the ordination scattergram. The Environmental Fuzzy Sets of A and Z have been both combined in a R-PCA (Orlóci 1978). The ordinations have been analyzed by the method of ellipses of equal concentration (Lagonegro and Feoli 1985a). Computations were carried out using the programs of Lagonegro and Feoli (1985b). Programs for computing fuzziness and anticommutative difference (FUZ and FUZZY) are available from V. Zuccarello.

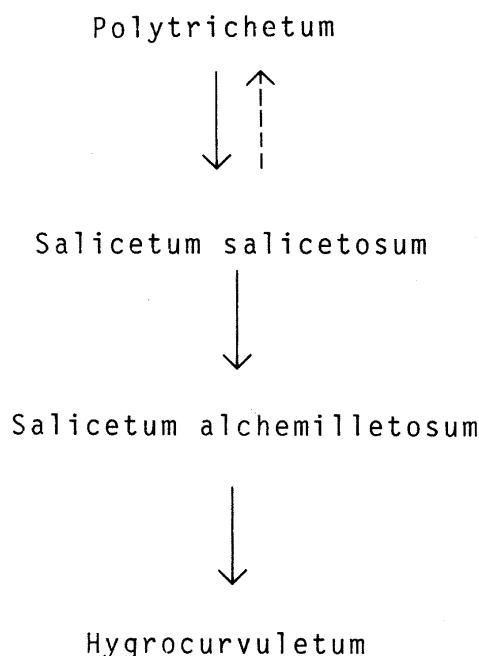


Fig. 1. A syndynamic scheme based on syntaxonomical data of alpine vegetation suggested by Giacomini and Pignatti (1955).

Fig. 2. Dendrogram of classification of the relevés of the 4 vegetation types in Fig. 1 and Table 1.

The classification of the 50 relevés is presented in the dendrogram of Fig. 2. The ordination based on fuzzy sets corresponding to type 1 and type 4 (*Polytrichetum* and *Hygrocurvuletum*), which are the most dissimilar types, is presented in Fig. 3a. The sequence suggested by the syndynamic scheme is retained on both axes. The distance between the centroids of the types is significant in all the cases, also between types 2 and 3 which have an apparent overlap. In this ordination, types 2 and 3, which are subassociations of the same association (*Salicetum herbaceae*), are apparently more closely related than the other types. The ordination

of the species of *Caricetea curvulae* and *Salicetea herbacea* according to the fuzzy sets corresponding to types 1 and 4 is presented in Fig. 3b. It may be overimposed on Fig. 3a in order to demonstrate the correspondence between types and species. The centroid of *Salicetea* species is placed just between the centroids of types 1 and 2, whereas the centroid of *Caricetea* species is placed between the centroids of types 3 and 4. The ellipsis of *Salicetea* shows an overlap with the ellipses of types 1 and 2; the ellipsis of *Caricetea* species shows an apparent overlap with the ellipses of types 3 and 4. The ordination shows that among the species of *Salicetea*, *Gymnomytrium varians*, *Pleuroclada albenscens*, *Anthelja juratzkana*, *Webera commutata*, *Webera carinata*, *Gentiana bavarica*, *Polytrichum sexangulare* are more related to *Polytrichetum*, whereas *Salix herbacea*, *Sedum alpestre* and *Cerastium cerastioides* are more related to *Salicetum salicetosum* and *Alchemilla pentaphyllea*, *Soldanella pusilla* and *Sibbaldia procumbens* are more related to *Salicetum alchemilletosum*. The ordination also shows that among the species of *Caricetea*, *Senecio carniolicus*, *Veronica bellidioides*, *Hieracium glanduliferum* and *Silene acaulis* are more related to *Hygrocurvuletum*, whereas *Ligusticum nuttellina* and *Potentilla aurea* are more related to *Salicetum alchemilletosum*. Furthermore *Minuartia sedoides* is more related to *Salicetum salicetosum*. The ordination pattern of species reproduces graphically the pattern of the structured table in Table 1.

The ordination of releves and types based on anti-commutative difference is presented in Fig. 4a, b. A

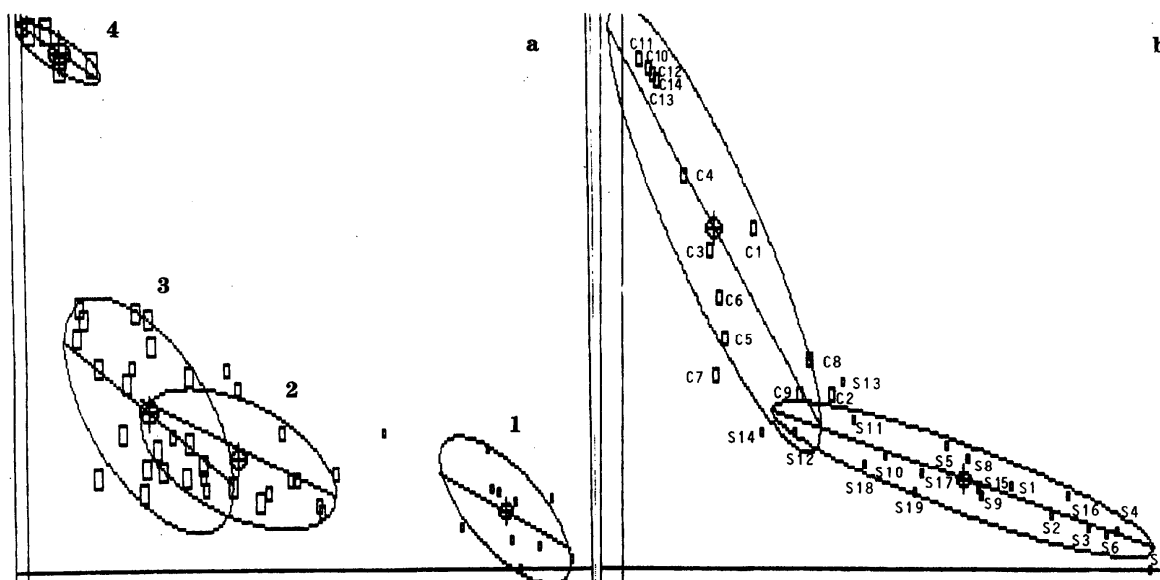


Fig. 3. Ordination of relevés and vegetation types (a) and species (b) of Salicetea (S) and Caricetea (C) on the basis of fuzzy sets corresponding to type 1 (abscissa) and type 4 (ordinate). Ellipses of equal concentration for probability 0.05. Numbers 1-4 identify types as in Fig. 1 and Table 1.

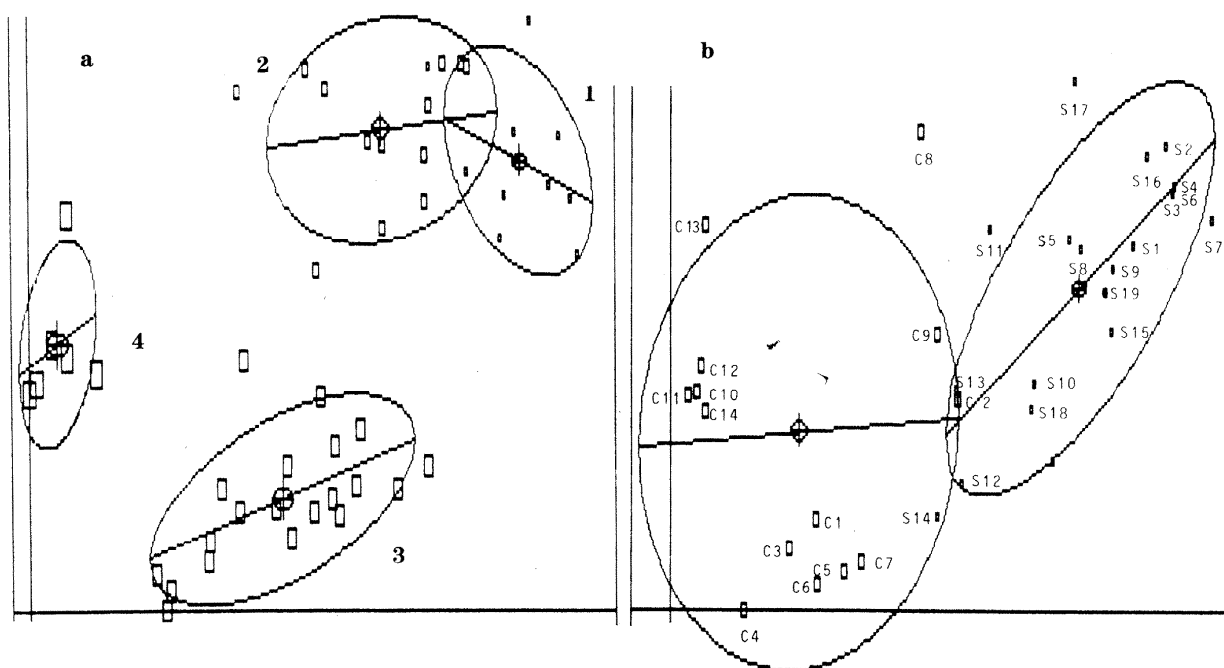


Fig. 4. Ordination of relevés and vegetation types (a) and species (b) of Salicetea (S) and Caricetea (C) by combining the fuzzy sets corresponding to types 1 and 4 (abscissa) and 2 and 3 (ordinate). Ellipses of equal concentration for probability 0.05. Numbers 1-4 identify the types as in Fig. 1 and Table 1.

clear separation of types 2 and 3 appears in Fig. 4a. The vertical axis is obtained by combining the fuzzy sets corresponding to these types. The sequence of the types in Fig. 1 is not retained along this axis because at its extremes only types 2 and 3 may be placed. This ordi-

nation offers a better resolving power of relevés and demonstrates the strict relationship between types 1 and 2 suggested by the double arrows in Fig. 1. Also in this example the ordination pattern of species (Fig. 4b) reproduces graphically the one in Table 1.

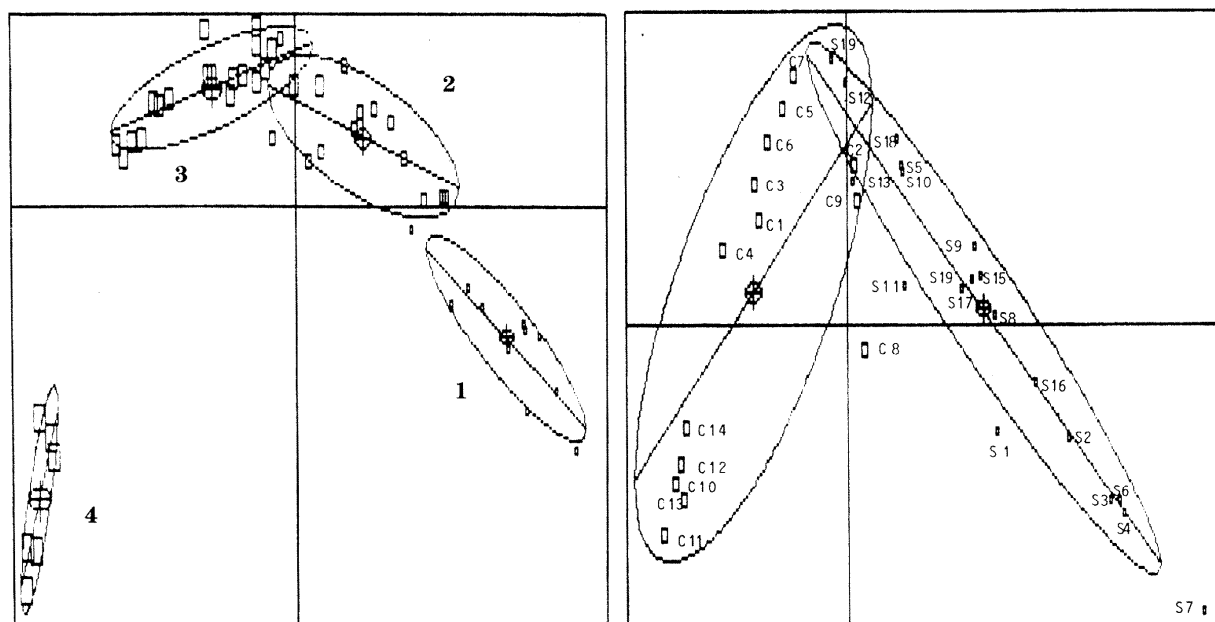


Fig. 5. Ordination of relevés and types (a) and species (b) of Salicetea (S) and Caricetea (C) according to PCA of environmental fuzzy sets. See the text for explanation. Ellipses of equal concentration for probability 0.05. Numbers 1-4 identify the types as in Fig. 1 and Table 1.

The ordinations based on environmental fuzzy sets (matrices **A** and **Z** combined in R-PCA) are given in Fig. 5 a and b. The correlation coefficients of EFS with the first components are given in Table 3. In both cases the first component explains a very high percentage of the total variance. Along the first component of Fig. 5a the sequence of the types follows that of Fig. 1. The centroids of the ellipses are all significantly separated. Fig. 5b shows the species of Salicetea well separated from those of Caricetea. Table 3 shows that the EFS are highly linearly correlated with the first two components in Fig. 5. From the correlation values of the EFS it follows that water content and organic matter are the environmental factors most related to the vegetational variation whereas pH is that least correlated.

Table 3. Correlation coefficients between the environmental variables and the first principal components (PCs) of the EFS matrices of relevés (rel.) and species (sp.).

| PC _s | pH | CaCO ₃ | P ₂ O ₅ t. | P ₂ O ₅ s. | O.M. | H ₂ O | EIGENVALUE | % |
|-----------------|-------|-------------------|----------------------------------|----------------------------------|--------|------------------|------------|-------|
| EFS rel. | 0.591 | 0.875 | -0.813 | -0.968 | -0.991 | -0.997 | 4.69 | 78.14 |
| EFS sp. | 0.893 | 0.981 | -0.829 | -0.973 | -0.982 | -0.996 | 5.35 | 89.19 |

Conclusion

The Braun Blanquet approach (Braun Blanquet 1964; Westhoff and van der Maarel 1978) has defined a very large numbers of vegetation types in different regions of the world. The typification involved visual arrangements of phytosociological tables and/or numerical methods of classification and ordination. In many papers visually-defined types have been confirmed by numerical methods (e.g. Moore 1972, Werger 1973, Feoli and Lagonegro 1982). The present paper shows the high correspondence between the visual classification of Giacomini and Pignatti (1955) and the results of a clustering method. The numerical analysis suggests that the three relevés (1, 2, 3) assigned to the subass. dicranetosum by Giacomini and Pignatti (1955), being well separated from the other relevés of Polytrichetum, should actually be assigned to the Anthelietum described by others (see Giacomini and Pignatti 1955).

The importance of vegetation types in ecological work has been recognized and discussed in many papers and books (e.g., Goodall 1963, Lieth 1968, Westhoff 1970, Whittaker 1970, Mueller-Dombois and Ellenberg 1974, van der Maarel and Werger 1978, Dierschke 1981). Personal opinions may differ (see McLean and Ivimey-Cook 1973, pp. 3431-3433), yet there is demonstrated need for hierarchical vegetation classifications. Ecological mapping at different scales and extrapolation of results obtained in the study of phytomass distribution and primary production could not be accomplished without such classifications. The present paper demonstrates on a simple data set that vegeta-

tion types can be used also as tools for explaining directly vegetation variation in relation to dynamics and gradients. They may be considered "one step in a further study" (Dale 1988) being the representatives of well defined clusters.

The use of syntaxonomical results in applications of fuzzy set theory reveals ordination patterns for species (variables) and relevés (syntaxonomical entities) along axes that are directly interpretable in terms of concepts or facts related to the biological composition of vegetation types, or in terms of environmental factors. This is an immense advantage over traditional ordination techniques and other methods based on eigenanalysis and non-metric multidimensional scaling (see Orlóci and Kenkel 1985). Another advantage is the possibility of carrying out direct ordination by environmental fuzzy sets notwithstanding the potential problem that environmental factors had not been measured in all the relevés. The approach based on fuzzy set theory shows that in an area where a syntaxonomical study is available, detailed ecological analysis on the relationships between vegetation types can be done by a very simple computational procedure.

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